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# Growth and Yield of Watermelon (*Citrullus lanatus*) in Plastic House in Response to White LED Supplementary Lighting

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## ABSTRACT

Watermelon plants cultivar 'Kinaree 457' were grown in plastic house under natural daylight only (control) or with nightly LED supplementary lighting for 6 h (6:00 pm-12:00 pm) or 12 h (6:00 pm-6:00 am) starting from transplanting up to fruit harvest. Plant height, leaf chlorophyll content and fruit yield significantly increased in response to 6 h supplementary LED lighting. Fruit mass, size (length x width) and flesh thickness at 6 h LED treatment were about 2.3 kg, 19.3 ×15.7 cm, and 15.7 cm, respectively, while the fruit of control had 1.7 kg, 16.0 × 14.3 cm, and 13.8 cm, respectively. No significant treatment effect was obtained on peel thickness, flesh color L\* and b\* values, juice pH and total soluble solids. However, 6 h LED treatment resulted in lower reddening flesh (lower a\* values), firmness and higher titratable acidity relative to the control, suggesting the need for improvement in cultural management. Furthermore, multivariate statistics of principal component analysis (PCA) performed on physico-chemical quality revealed the variations among watermelons from lighting and control treatments regardless of lighting hour.

# INTRODUCTION

Watermelon (*Citrullus lanatus*) is a popular cucurbit fruit-vegetable with high economic value and is widely grown all over the world (Wei et al., 2020) and in all regions of Thailand. Total production area in Thailand is about 13,740 hectares, with average yield of 21.1 tons per hectare and selling price of about 10.33 THB (0.3 USD) per kilogram (DOA, 2018). Watermelon is mainly consumed as fresh fruit or processed into food products such as watermelon juices and additives. It contains 34 kcal of energy, 92 g water, 0.7 g protein, 0.1 g fat and 7.0 g carbohydrates per 100 g fresh weight. Red-fleshed watermelons are rich in lycopene, which is higher than that in tomatoes. Market demand for watermelon continues to increase (Lignou, Parker,

Growing typical crops in greenhouses protects plants from harsh and unpredictable environments, insect pests and diseases infestations. In Thailand, the use of plastic house as greenhouse structure is now widespread due to its cost effectiveness and lengthy service life. Plastic houses are typically used for a wide variety of crops, particularly watermelons, cucumbers and melons, to obtain high quality products.

Light is the primary environmental factor that regulates plant growth and development (McNellis & Deng, 1995; Fukuda, Fujita, Ohta, Sase, Nishimura, & Ezura, 2008) as it is essential in photosynthetic and morphogenetic processes (Castellano et al., 2008). Three aspects of light affect plant growth - light intensity, light quality or wavelength, and

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light duration or photoperiod (Nadalini, Zucchi, & Andreotti, 2017), which can be controlled through artificial lighting to promote growth and increase quality yield of crops.

Light Emitting Diode (LED) is increasingly used to increase crop production efficiency in closed horticultural systems. The LED lamps usually last for 50,000-100,000 h compared to fluorescent lamps which last for only 2,000-3,000 hours. Furthermore, it consumes 81-90% less electricity than fluorescent lamps while produces at least 5 times more light than the conventional lamps and illuminates at least 2 times greater than Compact Fluorescent Lamps (CFL) (Mitchell et al, 2012). LED is also ecologically friendly as it does not emit infrared and ultraviolet rays which can be harmful to human skin and eyes and has no mercury component which makes it not destructive to the environment. LED is considered as a tool to promote and develop efficient plants (Folta, & Carvalho, 2015). It is used as a light source or as supplement to natural lighting (Li, & Kubota, 2009; Johkan, Shoji, Goto, Hashida, Yoshihara, 2010; Samuolienė, Sirtautas, Brazaityte, & Duchovskis, 2012; Choi, Moon, & Kang, 2015) usually under controlled conditions (Folta, & Carvalho, 2015; Yoshida, Hikosaka, Goto, Takasuna, & Kudou, 2012; Lin et al, 2013; Piovene et al, 2015). This study determined the efficacy of LED supplementary lighting in improving growth and fruit yield and quality of watermelon grown in plastic houses.

## MATERIALS AND METHODS

#### **Plant Samples**

Watermelon cultivar 'Kinaree 457' (Tawantonkla brand) was grown in 2019 in a 7 × 21 m<sup>2</sup> plastic house of the Faculty of Agricultural Technology, King Mongkut's Institute of Technology Ladkrabang (KMITL), Thailand. The plastic house was covered with 150  $\mu$ m-thick UV-treated polyethylene sheet. Seeds were sown in seedling tray and the plants with the same size were selected and thereafter transplanted individually into 8 × 15 inch<sup>2</sup> black plastic pots. The planting media was 3:1 garden soil:cow manure mixture. The pots were arranged in rows of 10 spaced at 0.4 × 0.6 m.

#### **Experimental Design**

A Completely Randomized Design (CRD) was used with the following treatments and every

treatment was replicated four times: natural daylight (control), natural daylight plus LED 6 h (6:00 pm – 12:00 pm) and natural daylight plus LED 12 h (6:00 pm-6:00 am). LED lighting was conducted using V-MAX 20W/6500K, size 100 × 100 × 170 mm (V-light brand, Five Lighting Co., Ltd., Thailand). The lighting was provided everyday starting from transplanting to harvest date.

#### **Cultural Management**

Fertilizer application, irrigation, weeding and insect pest and disease control followed commercial recommendations for plastic house-grown watermelons. The plants flowered after 20–25 days from transplanting and hand pollination was done to ensure fruit setting. One fruit was maintained per plant. At commercial maturity (about 50 days from fruit setting), the fruits were harvested for quality evaluation.

#### **Data Collection**

Plant height was determined using a metric rule (check) starting from 14 to 56 days after transplanting. Chlorophyll content of the 9th leaf from the apical tip was non-destructively measured using SPAD-502 chlorophyll meter (Minolta Camera Co., Japan) starting from 21 to 56 days after transplanting. At harvest, fruit were measured for weight using a digital weighing balance (Adventurer, OHAUS Corp., USA); width (equatorial diameter), length (polar diameter) and thickness of the peel and flesh using a Vernier caliper; color (L\*, a\*, b\*) using a Minolta Chromameter (CR-10); firmness using a penetrometer (Atago TR-53025, Italy); juice pH using a pH meter (Model HI2213, Hanna Instruments, USA); total soluble solids (TSS) using a refractometer (Model PAL-1, Tokyo, Japan); and titratable acidity (TA) by titration against a standard base (0.01N NaOH) using 1% phenolphthalein as indicator.

## **Statistical Analysis**

The results were analyzed by performing analysis of variance (ANOVA) and treatment means were analyzed based on Tukey's HSD test (p < 0.05) using Statistix 10 software.

Principal component analysis (PCA) was performed to preliminarily explore watermelon quality variation from all treatments using JMP version 10.0 software (SAS Institute Inc., Cary, NC, USA). The results were presented in score plot and biplot graphss.

## **RESULTS AND DISCUSSION**

#### **Plant Growth**

The plants increased in height more rapidly with LED lighting compared to that under natural daylight only (control) (Table 1). This effect was statistically significant after 28-56 days from transplanting. On the 56<sup>th</sup> and last day, plant height of the control (about 125 cm) was almost similar to that of LED-exposed plants after 28 days from transplanting. Final plant height of LED-treated plants was more than 10 cm longer than that of the control. The effects of 6 h and 12 h LED treatments were comparable, indicating that 6 h LED supplementary lighting was sufficient to promote growth.

Leaf chlorophyll content showed similar trend (Table 2). LED-treated plants had significantly higher chlorophyll content than the control after 21 days and after 42-56 days from transplanting. LED exposure duration did not show significant variations, thus 6 h LED lighting was enough for increasing chlorophyll content. The chlorophyll content decreased along with plant developmental stages. The chlorophyll content of control plant was recorded at 75 and 81-82 SPAD units at LED-treated plants after 21 days from transplanting and decreased to 58 (control) and 65-69 (LED-treated plants) SPAD units after 42-56 days from transplanting. One of the direct effects of LED supplementary lighting is the increase in plant height. This has been attributed to increased photosynthesis due to increased chlorophyll content. Both effects (increased plant height and chlorophyll content) were also obtained in the present study. As a result of increased photosynthesis, more organic substances such as sugars are produced, which support growth processes as source of energy (ATP) needed for metabolic processes and as carbon skeletons (precursors) of biosynthetic processes. Similar results were obtained in tomato (Watjanatepin, 2019), strawberry (Nadalini, Zucchi, & Andreotti, 2017), sweet pepper (Sobczak, Kowalczyk, Wolska, Kowalczyk, & Niedzi'nska, 2020) and melon (Kramchote, & Glahan, 2020).

#### Fruit Quality

LED lighting produced bigger and heavier fruits than the control (Table 3). LED exposures for 6 h and 12 h had comparable effect. The fruit weight of LEDtreated plants was 2.3-2.5 kg each and had 19-20 cm in length and 16 cm in width which were higher than that of the control (1.65 kg, 16 cm in length and 14 cm in width). Fig. 1 shows the size appearance of the fruits. In addition, fruits from LED-treated plants had thicker flesh (about 16 cm) than that of the control (14 cm) (Table 3). Peel thickness did not significantly vary among the treatments and ranged from 0.38 to 0.55 cm.

Treatmente	Days after transplanting								
Treatments	21	28	35	42	49	56			
No LED	75.23 <sup>⊳</sup>	72.05	77.13	64.68 <sup>b</sup>	62.28 <sup>⊳</sup>	58.25 <sup>⊳</sup>			
LED 6 h	80.60ª	77.23	75.13	71.83ª	68.20ª	64.73ª			
LED 12 h	81.70ª	79.15	68.48	74.60ª	71.48ª	69.13ª			
F-test	*	ns	ns	*	*	*			
C.V. (%)	3.96	5.90	6.02	5.85	5.49	6.06			

Table 1. Plant height (cm) of plastic house-grown watermelon in response to LED supplementary lightings

Remarks: Means in the same column followed by the same letter are not significantly different based on Tukey's HSD test; ns = not significant, \* = significant at 5%, \*\* = significant at 1%

Table 2.	Leaf ch	lorophyll	content	(SPAD	unit)	of plastic	house-grown	watermelon	in	response to	LED
suppleme	entary lig	ghting									

Treatments	Days after transplanting									
	14	21	28	35	42	49	56			
No LED	11.75	72.13	116.75 <sup>⊳</sup>	120.55 <sup>⊳</sup>	122.28 <sup>♭</sup>	123.50 <sup>b</sup>	124.83 <sup>₅</sup>			
LED 6 h	14.38	75.13	124.18ª	131.23ª	133.23ª	134.27ª	136.30ª			
LED 12 h	17.75	79.63	127.90ª	132.77ª	134.82ª	136.13ª	137.90ª			
F-test	ns	ns	**	**	**	**	**			
C.V. (%)	22.76	5.73	3.21	2.34	2.30	2.01	1.57			

Remarks: Means in the same column followed by the same letter are not significantly different based on Tukey's HSD test; ns = not significant, \* = significant at 5%, \*\* = significant at 1%

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Fruit flesh color coordinates L\* (lightness) and b\* (blue to yellow) did not differ statistically among treatments and ranged from 27.81-33.47 and 9.35-10.99, respectively (Table 4). Significant differences were obtained for a\* values (green to red) of fruit from 6 h LED treated plants which had lower a\* (15.66) indicating less red color than that the control (19.16). LED treatment for 12 h had a\* values comparable to the control. Fig. 2 shows the internal appearance of the fruit with the red flesh color appearing lighter in 6 h LED treatment compared to the other two treatments.

Firmness decreased in response to 6 h and 12 h LED treated plants from 12.69 to 7.66-7.90 N from the control (Table 4). Juice pH (5.52-6.87) and TSS ( $9.3-10.0^{\circ}B$ ) did not significantly differ among treatments while TA was significantly higher at 6 h LED treated plants (0.22%) relative to the control (0.15%) (Table 4).



Remarks: A = No LED, B = LED 6 h, C = LED 12 h

Fig. 1. External characteristics of plastic house-grown watermelons with or without LED supplementary lighting



Remarks: A = No LED, B = 6 h LED treatment, C = 12 h LED treatment

Fig. 2. Internal characteristics of plastic house-grown watermelons with or without LED supplementary lighting

**Table 3.** Fruit weight, length, width, peel thickness and flesh thickness of plastic house-grown watermelon in response to LED supplementary lighting

Treatments	Fruit weight (kg)	Fruit length (cm)	Fruit width (cm)	Peel thickness (cm)	Flesh thickness (cm)
No LED	1.65b	15.95b	14.29b	0.38	13.77b
LED 6 h	2.31a	19.25a	15.67ab	0.34	15.71a
LED 12 h	2.51a	20.47a	16.22a	0.55	16.38a
F-test	*	*	*	ns	*
C.V. (%)	17.74	9.42	5.68	53.31	7.27

Remarks: Means in the same column followed by the same letter are not significantly different based on Tukey's HSD test; ns = not significant, \* = significant at 5%, \*\* = significant at 1%

The results show that 6 h LED supplementary lighting was sufficient to increase fruit yield. The increase in fruit weight was due to the increase of fruit size that corresponded to the increase of flesh thickness but not peel thickness. This response to LED lighting supplement has been attributed to the increase of photosynthesis (Shimazaki, Doi, Assmann, & Kinoshita, 2007; Dong, Fu, Liu, & Liu, 2014; Sabzalian et al., 2014). The increase of photosynthetic capacity of LED-treated plants in the present study can be implicated from the increase of plant height and chlorophyll contents. This is in accordance with earlier findings in melon studies (Kramchote, & Glahan, 2020). The fruits produced with 6 h LED lighting were less red in flesh color implying lower lycopene content; softer (reduced firmness); and became more acidic (increased TA content). The sweetness was apparently not affected as TSS content was not changed relative to that of the control. Mechanisms underlying these changes can be examined in future studies.



**Fig. 3.** Score plot (A) of component 1 (PC1) against component 2 (PC2) of watermelon physico-chemical quality (L\*, a\*, b\*, firmness, pH, TSS and TA) obtained from all lighting treatments and biplot illustration (B) of PC1 versus PC2 of quality parameters are also presented

Treatments	Color						<b>TA</b> (0/)
	L*	a*	b*	Firmness (N)	рн	155 ( Drix)	IA (%)
No LED	30.96	19.16a	10.79	12.69a	5.69	9.33	0.15b
LED 6 h	27.81	15.66b	9.35	7.90b	5.52	10.00	0.22a
LED 12 h	33.47	16.66ab	10.99	7.66b	6.87	10.00	0.19ab
F-test	ns	*	ns	*	ns	ns	*
C.V. (%)	14.52	10.14	10.36	24.61	26.62	7.52	15.61

**Table 4.** Fruit flesh color, firmness, pH, total soluble solids (TSS) and titratable acidity (TA) of plastic housegrown watermelon in response to LED supplementary lighting

Remarks: Means in the same column followed by the same letter are not significantly different based on Tukey's HSD test; ns = not significant, \* = significant at 5%, \*\* = significant at 1%

## **Principal Component Analysis (PCA)**

A multivariate statistical analysis of PCA was applied to watermelons' physical and chemical quality in order to identify variance of each treatment and the quality parameter that dominantly affected by lighting condition. The study found that the set of two principal components (PC1 and PC2) accounted for 65.4% of total variance, and reached over 85.0% when considered together with PC3. Fig. 3 presents the PCA score plot of all studied parameters, illustrating two distinguishable clusters in the PCA space (Fig. 3A). The scenario suggested that LED lighting provided significant effects on both physical and chemical quality of watermelon regardless of lighting hour (6 and 12 hours). The PCA is often used to explore comprehensive variables and explain similarity or dissimilarity of samples (Kaewpangchan et al., 2021; Giuffrè, 2021). The study of Theanjumpol, & Maniwara, (2020) used PCA to explore quality variations of freshly harvested and 30-day stored pumpkins and discovered that both pumpkins had dissimilarity in most quality parameters especially for texture profiles. Likewise the aforementioned literature, in this study, quality traits from watermelons with no LED clearly showed dissimilarity with those from watermelons with LED exposure as they were clustered in the different side of the plane. Furthermore, according to bioplot graph (Fig. 3B), flesh color values (L\*, a\*, b\*) and firmness were considerably affected by LED lighting.

# CONCLUSION

LED supplementary lighting promoted growth and increased fruit yield of watermelons grown in plastic house. LED lighting duration of 6 h was sufficient as extending the exposure period to 12 h had no additional benefit. However, there were demerits of the treatment as the fruits had reduced reddening and firmness and increased acidity which suggest the need for further optimization studies to mitigate these effects.

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